

Follicular Variant of Papillary Thyroid Carcinoma: Cytological Indicators of Diagnostic Value

MANIMARAN D¹, KARTHIKEYAN T M², DOST MOHAMED KHAN³, THULASI RAMAN R⁴

ABSTRACT

Background: Fine needle aspiration cytology (FNAC) is an important investigation in preoperative diagnosis of thyroid lesions. Follicular variant of papillary carcinoma thyroid (FVPTC) is a well defined entity in histopathology, but its diagnosis in FNAC is usually missed and is challenging compared to classic papillary thyroid carcinoma.

Aims: The purpose of this study is to retrospectively analyze cytological features in histologically confirmed cases of FVPTC, compare them with literature and document the features that could increase the sensitivity of FNAC diagnosis.

Materials and Methods: Cytological smears from 22 histologically confirmed cases of FVPTC were evaluated for microscopic pattern and nuclear features by two independent pathologists and results compared with previous studies. Statistical analysis was done based on bivariate Pearson's correlation coefficient.

Results: Among 22 cases 21 were female and one male with age range 21 – 50 years. All patients had a solitary nodule except one with multicentric presentation. Preoperative cytological diagnosis were, classic papillary thyroid carcinoma (PTC); 7, FVPTC; 3, suspicious for PTC; 4, follicular neoplasm; 5 and adenomatous goiter; 3. Diagnosis upon cytological review were, FVPTC; 11, classic PTC; 7, suspicious for PTC; 2, follicular adenoma; 1 and adenomatous goiter; 1.

Conclusion: We conclude that cellular smears with features as observed in our case like microfollicular pattern, syncytial clusters, fine powdery chromatin, anisonucleosis and nucleomegaly should alert the pathologist to look carefully for other more specific features like nuclear grooves and nuclear pseudoinclusions. This approach will help in avoiding misdiagnosis of FVPTC and would aid in choosing the right treatment modality.

Keywords: Thyroid neoplasms, Papillary thyroid carcinoma, Fine needle aspiration cytology

INTRODUCTION

Papillary thyroid carcinoma (PTC) is the most common histological type of thyroid cancer and accounts for more than 80 % of papillary thyroid malignancies [1]. DeLellis defined papillary carcinoma as a well differentiated malignant epithelial tumor characterized by formation of papillary structures, psammoma bodies and a set of distinct nuclear features such as grooves, pseudoinclusions and ground glass appearance [2]. Many variants of papillary carcinoma have been described- oncocytic, tall cell, diffuse sclerosing, encapsulated and follicular variant which is the most common subtype [3]. Fine needle aspiration cytology (FNAC) is an important investigation in preoperative diagnosis of thyroid lesions [4]. Follicular variant of papillary thyroid carcinoma (FVPTC) is a well defined entity in histopathology, but its diagnosis in FNAC is usually missed and is challenging [5]. Preoperative diagnosis is clinically significant because FVPTC behaves similar to classic PTC and its misdiagnosis leads to conservatory surgery and a need for revision surgery after biopsy confirmation. Though many studies had been performed in this area, diagnosis of FVPTC in FNAC smears is still a challenging one. Our study analyzes the cytological features which would help to increase the sensitivity of FNAC in diagnosing this lesion with reasonable accuracy.

MATERIALS AND METHODS

Twenty six cases of histologically confirmed FVPTC with the FNAC slides were obtained from the pathology laboratory of a rural tertiary care referral hospital over a period of 6 years (2007 – 2012). Out of 26 cases, 4 were excluded from the study due to inadequate material for reporting. Smears stained with Papanicolaou and hematoxylin & eosin (H&E) stains were evaluated. Papanicolaou stained slides were analyzed because it increase the sensitivity of detection of nuclear features of PTC [6]. Cytomorphological features

were analyzed by two independent pathologists from a minimum two representative smears and the results were recorded. Presence of 5-6 groups of well-preserved follicular cells, with each group containing 10 or more cells on at least two slides from different passes satisfies the adequacy of the aspirate [7].

The features analyzed included cellularity, microscopic pattern (monolayered sheets, syncytial clusters, microfollicular pattern, papillary fronds and single cell distribution), nuclear features (anisonucleosis, nucleomegaly, fine powdery chromatin, grooves, intranuclear cytoplasmic inclusions, irregular nuclear membrane and nucleoli), colloid characteristics (thick or thin) and psammoma bodies. These features were graded as absent (0), occasional (1), frequently seen (2), prominent finding (3). Statistical analysis was done based on bivariate Pearson's correlation coefficient.

RESULTS

Among total cases, 21 were females and one was a male with age range 21 – 50 years. All patients had a solitary nodule of size 1 – 4.5 cm except for one patient with multicentric presentation. In the preoperative FNAC diagnosis, 14 cases were diagnosed with papillary carcinoma including classic PTC; 7, FVPTC; 3, PTC suspicious; 4. Diagnosis of remaining 8 cases revealed, follicular neoplasm; 5 and adenomatous goiter; 3. Diagnosis upon review were, FVPTC; 11, classic PTC; 7, PTC suspicious; 2, follicular neoplasm; 1 and adenomatous goiter; 1 [Table/Fig-1].

Our observation of nuclear features were shown in [Table/Fig-2]. Cases with moderate to high cellularity; 18 while low cellularity; 4 cases. Although multiple microscopic patterns in some cases were observed [Table/Fig-3-6], predominant microscopic pattern was microfollicular (20 cases), followed by syncytial clusters (17 cases). Grooves and pseudo inclusions were seen in 20 and 12 cases respectively [Table/Fig-7]. Fine powdery chromatin, anisonucleosis

Diagnostic category	Number of cases	
	Preoperative diagnosis	After review
Classic PTC	7	7
FVPTC	3	11
Suspicious of PTC	4	2
Follicular neoplasm	5	1
Adenomatous goiter	3	1
Total	22	22

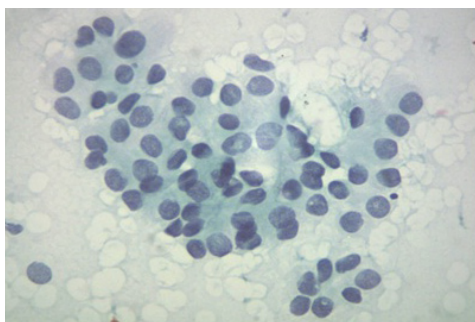
[Table/Fig-1]: Preoperative diagnosis by FNAC and after review PTC – papillary thyroid carcinoma, FVPTC- follicular variant of papillary thyroid carcinoma

Microscopic pattern				
Grading	0 – absent	1- occasional	2 – frequent	3 – prominent
Microfollicular pattern	2	2	12	6
Syncytial clusters	5	4	10	3
Monolayered sheets	10	2	6	4
Papillary fronds	15	3	3	1
Single cells	14	3	4	1
Nuclear features				
Nuclear grooves	2	6	9	5
Pseudoinclusion	12	3	5	2
Fine powdery chromatin	0	3	8	11
Anisonucleosis	0	1	3	18
Nucleomegaly (2 times RBC)	0	1	2	19
Irregular nuclear membrane	10	6	4	2
Nucleoli	9	8	4	1

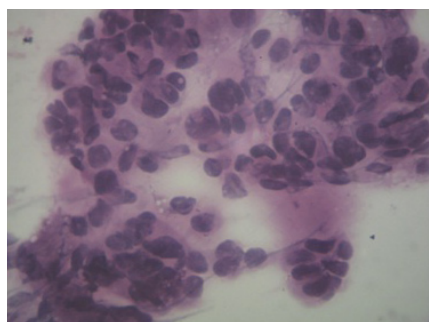
[Table/Fig-2]: Microscopic pattern and nuclear features

and nucleomegaly were seen in all cases [Table/Fig-8]. Number of cases with thick chewing gum colloid [Table/Fig-9]; 16, hurthle cells; 5, and psammoma bodies not identified in any of these cases.

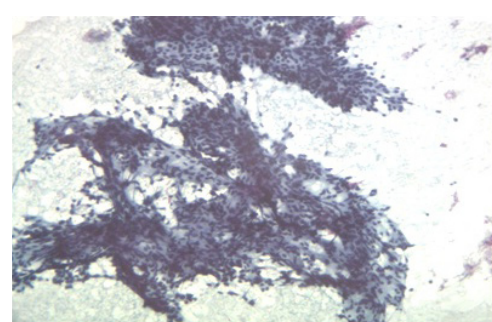
The p-value for microfollicular pattern, monolayered sheets, papillary fronds, nuclear grooves, nuclear pseudoinclusions and nucleoli were found to be significant.



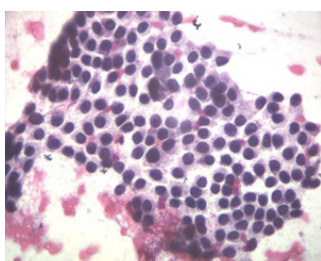
[Table/Fig-3]: Cells in microfollicular pattern. (Papanicolaou stain, x400)



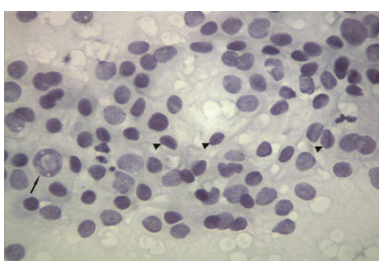
[Table/Fig-4]: Cells in syncytial clusters. (H&E stain, x400)



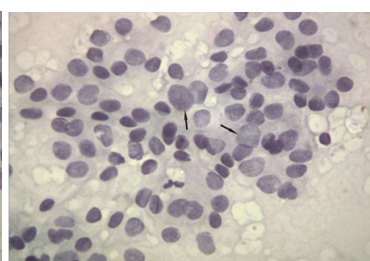
[Table/Fig-5]: Cells in papillary fronds. (Papanicolaou stain, x40)



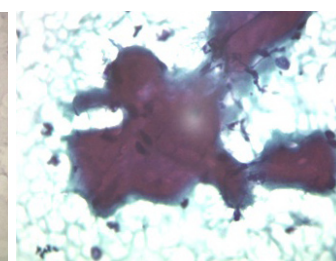
[Table/Fig-6]: Cells in monolayered sheets. (H&E stain, x100)



[Table/Fig-7]: show intranuclear pseudoinclusion (arrow) and nuclear grooves (arrow heads). (Papanicolaou stain, x400)



[Table/Fig-8]: show anisonucleosis, nucleomegaly (arrow) and fine powdery chromatin (Papanicolaou stain, x400)



[Table/Fig-9]: Thick chewing gum colloid (Papanicolaou stain, x400)

DISCUSSION

FVPTC was first described by Crile and Hazard who named this lesion alveolar variant of PTC [8]. This was confirmed by Lindsay and the author observed that although the neoplasm had a follicular architecture, the nuclear features were that of conventional PTC. Therefore the tumor should be designated FVPTC. Chen and Rosai stressed the importance of nuclear rather than architectural pattern in making a diagnosis of PTC and showed that FVPTC behaved similarly to the conventional PTC [9].

Because of the follicular pattern it was often confused with other benign and neoplastic follicular lesions including adenomatoid nodule, follicular adenoma and follicular carcinoma [10,11]. The diagnosis was solely based on nuclear features and owing to paucity of these findings on aspiration material, some of these lesions could be easily missed [7]. Even focal nuclear features of papillary carcinoma thyroid in fine needle aspiration cytology are strongly associated with papillary carcinoma at resection, [12] particularly FVPTC [13]. Higher tumor size and rate of suspicious for malignancy cytology were significantly higher in FVPTC [14]. If we use histologic diagnosis as the gold standard, the sensitivity and specificity of FNAC diagnosis of FVPTC were 42% and 83%, respectively [15].

In a study by Lin HS et al., [2] sensitivity of FNAC for the diagnosis of FVPTC was only 25% compared to 74% for the classic PTC. There are two reasons for the low sensitivity of FNAC in its diagnosis. One is FVPTC has features in common with benign and neoplastic follicular lesions because of the presence of colloid and monolayered sheets of follicular cells. Second, FVPTC show very few nuclear features of classic PTC. Most of the cases in our study were initially misdiagnosed as follicular neoplasm and adenomatous goiter. This may lead to inadequate surgical treatment like lobectomy and isthmusectomy. No well-defined minimal cytological criteria for the diagnosis of FVPTC in FNAC has been established [4]. This underscores the significance of increasing the accuracy of FNAC in the diagnosis of FVPTC.

Aron et al., [16] in their study stressed that syncytial clusters, microfollicular pattern, chromatin clearing and nuclear grooves were significant features to the diagnosis of FVPTC. Nucleomegaly was seen in all of their cases. In a study by Wu et al., [4] flat

syncytial sheets, nuclear enlargement and fine chromatin were seen in all cases of FVPTC along with thick colloid in 20/29 cases. These syncytial sheets showed branching with nuclear crowding compared to honeycomb pattern of colloid nodule. In our study microfollicular and syncytial clusters were the predominant patterns while thick colloid was seen in 16 cases. Fine powdery chromatin, anisonucleosis and nucleomegaly were seen in all of our cases.

According to Powari et al., [17] FVPTC should be considered in the diagnosis if the smears show high cellularity, syncytial clusters and follicular arrangement and thick colloid. Shih SR et al., [15] concluded in their study that it was difficult to differentiate FVPTC from classic PTC in FNAC and when follicular structures were seen in smears, careful search for nuclear features should be done. In the present study papillary structures were seen in 7 cases all were diagnosed as classic PTC even after review.

In a study by Gallagher et al., [8] most of the smears were highly cellular and the neoplastic cells formed follicles, tubules, rosettes, papillary structures and the nuclei were twice the size of red blood cells, hyperchromatic and had smooth contours. Yan et al., [18] in their study showed that branched sheets of follicular cells seen in low-power examination of Diff-Quick-stained smears are clue to the quick diagnosis of FVPTC and the diagnosis can be confirmed by further evaluation of nuclei in Papanicolaou-stained smears. In our study we analysed papanicolaou stained slides along with H&E slides and identified nuclear grooves in 20 cases, pseudoinclusions in 12 cases.

We conclude that cellular smears with features as observed in our case like microfollicular pattern, syncytial clusters, fine powdery chromatin, anisonucleosis and nucleomegaly should alert the pathologist to look carefully for other more specific features like nuclear grooves and nuclear pseudoinclusions. This approach will help in avoiding misdiagnosis of FVPTC and would aid in choosing the right treatment modality.

REFERENCES

- [1] Baloch ZW, LiVolsi VA. Pathology of thyroid and parathyroid disease. In: Mills SE, editor. Strenberg's diagnostic surgical pathology. 5th ed. Virginia: Lippincott Williams & Wilkins; 2010. p. 499.
- [2] Lin HS, Komisar A, Opher E, Blaugrund SM. Follicular Variant of Papillary Carcinoma: The Diagnostic Limitations of Preoperative Fine-Needle Aspiration and Intraoperative Frozen Section Evaluation. *Laryngoscope*. 2000 Sep;110(9):1431-6.
- [3] Zidan J, Karen D, Stein M, Rosenblatt E, Basher W, Kuten A. Pure versus Follicular Variant of Papillary Thyroid Carcinoma : Clinical Features, Prognostic Factors, Treatment, and Survival. *Cancer*. 2003 Mar 1;97(5):1181-5.
- [4] Wu HH, Jones JN, Grzybicki DM, Elsheikh TM. Sensitive Cytologic Criteria for the Identification of Follicular Variant of Papillary Thyroid Carcinoma in Fine-Needle Aspiration Biopsy. *Diagn Cytopathol*. 2003 Nov;29(5):262-6.
- [5] Martínez-Parra D, Campos Fernández J, Hierro-Guilmain CC, Sola Pérez J, Pérez-Guillermo M. Follicular variant of papillary carcinoma of the thyroid: to what extent is fine-needle aspiration reliable?. *Diagn Cytopathol*. 1996 Jul;15(1):12-6.
- [6] Yang GC, Liebeskind D, Messina AV. Diagnostic accuracy of follicular variant of papillary thyroid carcinoma in fine-needle aspirates processed by ultrafast Papanicolaou stain: histologic follow-up of 125 cases. *Cancer*. 2006 Jun 25;108(3):174-9.
- [7] Bommanahalli BP, Bhat RV, Rupanarayan R. A cell pattern approach to interpretation of fine needle aspiration cytology of thyroid lesions: A cyto-histomorphological study. *J Cytol*. 2010 Oct;27(4):127-32.
- [8] Gallagher J, Oertel YC, Oertel JE. Follicular variant of papillary carcinoma of the thyroid: fine-needle aspirates with histologic correlation. *Diagn Cytopathol*. 1997 Mar;16(3):207-13.
- [9] Salajegheh A, Petcu EB, Smith RA, Lam AK. Follicular variant of papillary thyroid carcinoma: a diagnostic challenge for clinicians and pathologists. *Postgrad Med J*. 2008 Feb;84(988):78-82.
- [10] Deveci MS, Deveci G, LiVolsi VA, Baloch ZW. Fine-needle aspiration of follicular lesions of the thyroid. Diagnosis and follow-Up. *Cytojournal*. 2006 Apr 7;3:9.
- [11] Baloch ZW, LiVolsi VA. Encapsulated Follicular Variant of Papillary Thyroid Carcinoma with Bone Metastases. *Mod Pathol*. 2000 Aug;13(8):861-5.
- [12] Renshaw AA. Focal Features of Papillary Carcinoma of the Thyroid in Fine-Needle Aspiration Material Are Strongly Associated With Papillary Carcinoma at Resection. *Am J Clin Pathol*. 2002 Aug;118(2):208-10.
- [13] Logani S, Gupta PK, LiVolsi VA, Mandel S, Baloch ZW. Thyroid nodules with FNA cytology suspicious for follicular variant of papillary thyroid carcinoma: follow-up and management. *Diagn Cytopathol*. 2000 Dec;23(6):380-5.
- [14] Ozdemir D, Ersoy R, Cuhaci N, Arpacı D, Ersoy EP, Korukluoglu B, et al. Classical and follicular variant papillary thyroid carcinoma: comparison of clinical, ultrasonographical, cytological, and histopathological features in 444 patients. *Endocr Pathol*. 2011 Jun;22(2):58-65.
- [15] Shih SR, Shun CT, Su DH, Hsiao YL, Chang TC. Follicular variant of papillary thyroid carcinoma: diagnostic limitations of fine needle aspiration cytology. *Acta Cytol*. 2005 Jul-Aug;49(4):383-6.
- [16] Aron M, Mallik A, Verma K. Fine needle aspiration cytology of follicular variant of papillary carcinoma of the thyroid , morphological pointers to its diagnosis. *Acta Cytol*. 2006 Nov-Dec;50(6):663-8.
- [17] Powari M, Dey P, Saikia UN. Fine needle aspiration cytology of follicular variant of papillary carcinoma of thyroid. *Cytopathology*. 2003 Aug;14(4):212-5.
- [18] Yan Z, Yang GC, Waisman J. A low-power, "architectural," clue to the follicular variant of papillary thyroid adenocarcinoma in aspiration biopsy. *Acta Cytol*. 2000 Mar-Apr;44(2):211-7.

PARTICULARS OF CONTRIBUTORS:

1. Associate Professor, Department of Pathology, Shri Sathya Sai Medical College and Research Institute, Tiruporur, Tamilnadu, India.
2. Associate Professor, Department of Pathology, Melmaruvathur Adhiparasakthi Institute of Medical Science and Research, Melmaruvathur, Tamil Nadu, India.
3. Assistant Professor, Department of Pathology, Shri Sathya Sai Medical College and Research Institute, Tiruporur, Tamilnadu, India.
4. Assistant Professor, Department of Pathology, Shri Sathya Sai Medical College and Research Institute, Tiruporur, Tamilnadu, India.

NAME, ADDRESS, E-MAIL ID OF THE CORRESPONDING AUTHOR:

Dr. Manimaran D,
New no 5, LIC Colony Main Road, Velachery, Chennai, Tamilnadu- 600042, India.
Phone: 9841310232, E-mail: manimaran.anu@gmail.com

FINANCIAL OR OTHER COMPETING INTERESTS: None.

Date of Submission: **Aug 26, 2013**
Date of Peer Review: **Dec 08, 2013**
Date of Acceptance: **Jan 13, 2014**
Date of Publishing: **Mar 15, 2014**